Introduction

Atopic dermatitis (AD) is a chronic inflammatory skin disease with a high prevalence in children. AD is caused by a combination of various causes. Patients with atopic dermatitis complain of symptoms such as erythema, edema, severe itching, and exudation. Atopic dermatitis patients often have allergic rhinitis and asthma. These diseases progress chronically due to environmental changes due to industrialization, improper eating habits, and stress, and continue into adulthood.

In atopic dermatitis, the skin is dry or itchy, and eczema erythematous appears. These skin symptoms are due to damage to the skin barrier. When ceramide is reduced in the stratum...
corneum of the skin, the skin barrier function and moisture retention function that ceramide is responsible for decreases\(^5\). The causes of atopic dermatitis are diverse and complex. Physical and functional damage of the skin barrier occurs due to various causes, resulting in secondary immune system problems\(^6\).

In Korean traditional medicine, atopic dermatitis is caused by moist heat (濕熱) caused by congenital weakness, or by the intrusion of wind heat (風熱) from the outside and the accumulation of moist heat inside\(^7\). Treatments includes ‘dispel wind to clear heat (祛風清熱)’ and ‘nourish blood to moisten dryness (養血潤燥)’\(^8\). Therefore, in the clinical field, herbs with a clear heat (清熱) effect are often used for the treatment of atopic dermatitis\(^9\).

*Coptis chinensis* (黃連) has medicinal effects of ‘clear heat and dry dampness (清熱燥濕)’ and ‘purge fire and detoxify (瀉火解毒)’, and also has antiviral and antibacterial effects\(^10,11\). According to previous study\(^12\), *Coptis chinensis* is also effective in suppressing inflammation and repairing skin damage from burns. Therefore, we tried to confirm the possibility of atopic dermatitis treatment by inhibiting skin damage and restoring the damaged skin barrier of *Coptis chinensis* extract-ceramide complex.

This study confirmed the regulation of the Endocannabinoid system (ECS) through cannabinoid receptor (CB) 1 and CB2. In addition, the regulation of macrophage activity through changes in Cluster of Differentiation 68 (CD68), phosphorylated inhibitor kappa B (p-I\(_\kappa\)B), and inducible nitric oxide synthase (iNOS) and control of mast cell activity through changes in substance P and serotonin were confirmed. Through this, we report the possibility that *Coptis chinensis* extract-ceramide complex can alleviate the inflammatory response of atopic dermatitis.

### Materials and Methods

#### 1. Materials

1) Animals

As the experimental animals, 6-week-old male Balb/C mice (20 ± 1.5 g, Jabio, Suwon, Korea) selected after acclimatization for 2 weeks in an aseptic breeding apparatus were used. They were divided into normal group (Ctrl), lipid barrier elimination group (ADE), palmitoylethanolamide (PEA) treated group after lipid barrier elimination (PEAT), and *Coptis chinensis* extract-ceramide complex applied group after lipid barrier elimination (CRA). 10 animals were assigned to each group. Animal experiments were conducted after approval from the Institutional Animal Care and Use Committee of Pusan University (IACUC No. PNU-2015-0924). The care and use of laboratory animals was conducted in accordance with NIH guidelines.

2) Experimental drug

*Coptis chinensis* (50 g, Omniherb, Daegu, Korea) was added in 100 ml of distilled water, boiled for 3 hours, and then filtered. It was reduced to 50 ml using a rotary evaporator, concentrated and freeze-dried to extract 8.2 g (yield: 16.4%). This extract was diluted with physiological saline to make a 5% extract dilution. This dilution was mixed with 5% ceramide 3B (Ecofactory, Incheon, Korea) in a 1:1 ratio to...
make a 5% *Coptis chinensis* extract-ceramide complex.

To CRA, 100 μl of a 5% *Coptis chinensis* extract-ceramide complex was applied to the lipid barrier eliminated skin for 5 days. To PEAT, PEA (Sigma-Aldrich, St. Louis, MO) used as a control drug was dissolved in physiological saline (100 μl) and then 10 mg/kg was orally administered for 5 days after eliminating the lipid barrier.

### 2. Methods

#### 1) lipid barrier elimination

To eliminate the lipid barrier, the dorsal skin of the mouse was shaved, and then the stratum corneum desquamation was removed using Tape (3M, St. Paul, MN). 500 μl of 10% sodium dodecyl sulfate (SDS: Sigma-Aldrich, St. Louis, MO) was applied on the spot and then rubbed 30 times using a cotton swab to remove lipid lamella from the stratum corneum.

#### 2) Tissue section production

Cardiac perfusion fixation was performed on the skin with vascular rinse and 10% neutral buffered formalin (NBF). After the obtained dorsal skin was fixed in 10% NBF for 24 hours, it was embedded in paraffin by a conventional method, and serial sections were made with a thickness of 5 μm.

#### 3) Immunohistochemical test

Immunohistochemical staining was performed to investigate the immunohistological changes of the ECS, macrophage activity and mast cell activity. For immunohistochemical staining, antibodies such as anti-CB1, anti-CB2, anti-CD68, anti-p-IκB, anti-Substance P and anti-Serotonin were used.

First, the skin sections were subjected to proteolysis in proteinase K (20 μg / ml, Agilent Dako, Santa Clara, CA) for 5 minutes and then treated with 10% normal goat serum (Vector Lab, Burlingame, CA) containing 1% fetal bovine serum (Sigma-Aldrich, St. Louis, MO) for 1 hour. Then, the primary antibody, mouse anti-CB1 (1:100, Santa Cruz Biotec, Dallas, TX), mouse anti-CB2 (1:50, Santa Cruz Biotec), mouse anti-CD68 (1:50, Santa Cruz Biotec), mouse anti-p-IκB (1:500, Abcam, Cambridge, UK), mouse anti-iNOS (1:250, Abcam), mouse anti-substans P (1:100, Santa Cruz Biotec) and mouse anti-serotonin (1:50, Santa Cruz Biotec) were reacted in a humidified chamber at 4 °C for 72 hours. The secondary antibody, biotinylated goat anti-mouse IgG (1:100, Abcam), was linked for 24 hours at room temperature, and reacted with avidin biotin complex kit (Vector Lab) for 1 hour at room temperature. After color development in 0.05 M tris-HCl buffer (pH 7.4) containing 0.05% 3,3′-diaminobenzidine and 0.01% HCl, counterstaining was performed with hematoxylin.

#### 4) Image analysis

The results of immunohistochemistry were quantified (means ±standard error) through image analysis using image Pro 10 (Media cybernetics, Rockville, MD). After randomly selecting 10 skin samples from each group, they were photographed at x400 magnification, and then images were analyzed with positive pixels (intensity 80-100) /20,000,000 pixels.
5) Statistics

For statistics, SPSS software (SPSS 25, SPSS Inc., Chicago, IL) was used. Significance \( p < 0.05 \) was verified through one-way ANOVA, and Tukey HSD was performed for post hoc verification.

Results

1. ECS regulation

CB1 positive reaction was increased in the ADE, PEAT, and CRA compared to the Ctrl (6,730 ± 390 /20,000,000 pixel). The ADE (48,832 ±1,716 /20,000,000 pixel) increased by 626% compared to the Ctrl. The PEAT (59,878 ± 6,161 /20,000,000 pixel) increased by 790% compared to the Ctrl, and the CRA (72,024 ± 2,471 /20,000,000 pixel) increased by 970% compared to the Ctrl. The CRA had significantly a 48% higher positive reaction than the ADE and 20% than the PEAT (Fig. 1).

CB2 positive reaction was increased in the ADE, PEAT, and CRA compared to the Ctrl (6,583 ± 274 /20,000,000 pixel). The ADE (32,986 ± 1,653 /20,000,000 pixel) increased by 401% compared to the Ctrl. The PEAT (44,704 ± 965 /20,000,000 pixel) increased by 579% compared to the Ctrl, and the CRA (58,739 ± 992 /20,000,000 pixel) increased by 792% compared to the Ctrl. The CRA had significantly a 78% higher positive reaction than the ADE and 31% than the PEAT (Fig. 1).

2. Macrophage activity regulation

CD68 positive reaction was increased in the ADE, PEAT, and CRA compared to the Ctrl (6,730 ± 390 /20,000,000 pixel). The ADE (48,936 ± 1,716 /20,000,000 pixel) increased by 626% compared to the Ctrl. The PEAT (59,878 ± 6,161 /20,000,000 pixel) increased by 790% compared to the Ctrl, and the CRA (72,024 ± 2,471 /20,000,000 pixel) increased by 970% compared to the Ctrl. The CRA had significantly a 48% higher positive reaction than the ADE and 20% than the PEAT (Fig. 1).
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Compared to the Ctrl. The PEAT (47,419 ± 1,205/20,000,000 pixel) increased by 532% compared to the Ctrl, and the CRA (37,354 ± 2,672/20,000,000 pixel) increased by 398% compared to the Ctrl. The CRA had significantly a 54% lower positive reaction than the ADE and 21% than the PEAT (Fig. 2).

p-IκB positive reaction was increased in the ADE, PEAT, and CRA compared to the Ctrl (6,147 ± 370/20,000,000 pixel). The ADE (80,073 ± 1,889/20,000,000 pixel) increased by 1,203% compared to the Ctrl. The PEAT (44,090 ± 1,429/20,000,000 pixel) increased by 617% compared to the Ctrl. The CRA (29,703 ± 2,351/20,000,000 pixel) increased by 383% compared to the Ctrl. The CRA had significantly a 63% lower positive reaction than the ADE and 33% than the PEAT (Fig. 2).

iNOS positive reaction was increased in the ADE, PEAT, and CRA compared to the Ctrl (6,068 ± 420/20,000,000 pixel). The ADE (60,145 ± 1,701/20,000,000 pixel) increased by 891% compared to the Ctrl. The PEAT (36,406 ± 1,335/20,000,000 pixel) increased by 500% compared to the Ctrl, and the CRA (21,240 ± 673/20,000,000 pixel) increased by 250% compared to the Ctrl. The CRA had significantly a 65% lower positive reaction than the ADE and 42% than the PEAT (Fig. 2).

3. Mast cell activity regulation

Substance P positive reaction was increased in the ADE, PEAT, and CRA compared to the Ctrl (5,088 ± 246/20,000,000 pixel). The ADE (74,641 ± 1,663/20,000,000 pixel) increased by 1367% compared to the Ctrl. The PEAT (57,047 ± 1,861/20,000,000 pixel) increased by 1,021% compared to the Ctrl, and the CRA (25,610 ± 854/20,000,000 pixel) increased by 1,021% compared to the Ctrl, and the CRA (25,610 ± 854/20,000,000 pixel).
The positive reaction of Substance P and serotonin (arrow indicates light brown particle) is higher in the ADE, PEAT and CRA compared to the Ctrl, but the increase in the positive reaction was lower in the CRA compared to the ADE and PEAT. *, p < 0.05 compared with ADE; #, p < 0.05 compared with PEAT. Abbreviation same as Fig. 1.

Atopic dermatitis is a chronic disease that deteriorates the quality of life due to recurrence and worsening of symptoms at various ages. There are various causes of atopic dermatitis, and at present, there is no fundamental treatment, only to relieve symptoms or prevent further deterioration. Currently, antihistamines and steroids are used for the treatment of atopic dermatitis, but it is only a temporary symptom relief rather than a fundamental treatment.

Coptis chinensis extract-ceramide complex suppresses inflammation and restores skin damage with its cleansing heat and detoxifying effect. In this study, we confirmed the possibility that Coptis chinensis extract-ceramide complex could be a therapeutic agent for atopic dermatitis by inhibiting skin damage and restoring damaged skin barrier.

The ECS is a biological system that regulates appetite, pain sensation, mood, and memory. The
ECS consists of cannabinoid receptors (CBRs), endogenous ligands of CBRs, and enzymes involved in the synthesis and degradation of CB1. Increased or decreased the ECS has been associated with various pathological conditions. Transient changes in ECS action can reduce or slow the progression of various bodily compensatory responses or diseases. On the other hand, the activity of the ECS may reflect bodily defects or act pathogenic. In particular, the ECS is involved in various responses of skin cells. Activation of CB1 and CB2 in epidermal cells inhibits keratinocyte proliferation and differentiation, induces inflammation, and induces apoptosis. Various CBs suppress sensory symptoms such as pain and itching through CB1. The ECS regulates immune and inflammatory responses as well as cell differentiation in the skin. Therefore, even fine adjustment of the ECS can promote or alleviate skin diseases such as atopic dermatitis.

For this reason, research using the ECS as a new treatment for skin diseases is emerging.

In this study, both PEAT and CRA increased significantly in CB1 and CB2 than ADE, and CRA increased more than PEAT. The results of this study indicate that Coptis chinensis extract-ceramide complex produces the ECS components such as CB1 and CB2. This suggests the possibility of alleviating the symptoms of atopic dermatitis through the intervention of the Coptis chinensis extract-ceramide complex in the action of the ECS.

Mitogen-activated protein kinases (MAPKs), including extracellular signal-regulated kinase (ERK) and c-Jun N-terminal kinases (JNK), regulate the inflammatory response through nuclear factor kappa B (NF-κB) activation. NF-kB is a key regulator in the cellular inflammatory response and exists in an inactive state in the cytoplasm in a state of binding with inhibitor of NF-kB (IκB). When a cell is damaged or stressed, NF-kB is separated into IκB by IκB kinase (IKK), and enters the nucleus to cause an inflammatory response. And iNOS, which is transcribed by the pro-inflammatory enzyme produced by NF-kB, excessively produces nitric oxide (NO) to create oxidative stress, thus causing tissue damage and inflammation.

In this study, both PEAT and CRA decreased significantly in CD68, p-IκB, and iNOS than ADE, and CRA decreased more than PEAT. This means that the Coptis chinensis extract-ceramide complex is likely to be involved in the regulation of macrophage activity.

Symptoms of atopic dermatitis, such as severe pruritus, are initiated by immunoglobulin E (IgE) -mediated degranulation of mast cells. When IgE and antigen bind to receptors expressed on the surface of mast cells, mast cells are activated, and the activated mast cells secrete inflammatory mediators such as substance P and Matrix metalloproteinase (MMP) -9 to induce an inflammatory response. Substance P not only regulates the secretion of various cytokines, but also participates in the inflammatory response and induces itching through vascular relaxation, increased vascular permeability, and histamin secretion. So, Substance P can be said to be an indicator of itching symptoms. Serotonin exists in platelets, activates dermal mast cells, secretes histamine, and acts on 5-HT3 receptors in the central nervous system to induce pruritus.
In this study, substance P and serotonin showed higher positive reaction in the ADE, PEAT and CRA than in the Ctrl, but the increase in the positive reaction was lower in the CEA compared to the ADE and PEAT. This result means that the *Coptis chinensis* extract-ceramide complex can modulate mast cell activity. This shows the possibility that the *Coptis chinensis* extract-ceramide complex can alleviate symptoms of atopic dermatitis such as itching.

*Coptis chinensis* extract-ceramide complex inhibited the pathological symptoms of the skin barrier and skin damage in atopic dermatitis. In addition, it promoted the formation of the ECS components such as CB1 and CB2 and a lipid barrier, and inhibited proliferation and invasion of inflammatory cells.

These results show that *Coptis chinensis* extract-ceramide complex maintains skin homeostasis, improves skin barrier formation and suppresses inflammation through the ECS intervention. Therefore, *Coptis chinensis* extract-ceramide complex is expected to alleviate the symptoms of atopic dermatitis. However, since this study was limited to animal experiments, additional clinical studies are needed in the future.

**Conclusion**

In this study, the effects of *Coptis chinensis* extract-ceramide complex were confirmed on the regulation of the ECS through CB1 and CB2, the regulation of macrophage activity through CD68, p-1κB, and iNOS, and the regulation of mast cell activity through substance P and serotonin.

1. CB1 and CB2 showed higher positive reactions in the CRA than in the ADE and PEAT.
2. CD68, p-1κB and iNOS showed higher positive reaction in the ADE, PEAT and CRA than in the Ctrl, but the increase in the positive reaction was lower in the CEA compared to the ADE and PEAT.
3. Substance P and serotonin showed higher positive reaction in the ADE, PEAT and CRA than in the Ctrl, but the increase in the positive reaction was lower in the CEA compared to the ADE and PEAT.

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**References**

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ORCID

Sang Hyun Ahn  https://orcid.org/0000-0002-1526-4397
Ki Bong Kim  https://orcid.org/0000-0002-0096-5836